1. Procedure Description
   Include exact details of any/all chemical, biological, radiation, or physical agents as well as route(s)/dose(s)/volume(s)/frequency and duration

Method 1:
1. Restrain the animal by gently grasping the base of the tail with dominant hand. With non-dominant hand, place thumb and index finger, or second knuckle, on either side of the neck and draw up the loose skin, trapping the fold gently between the finger and thumb to ensure that the weight of the animal is supported. The tail may then be released or restrained.

   Alternative method of restraint (one-handed method):
   Place the tail of the animal between two fingers of the non-dominant hand or secure tail with pinkie finger. Place the same hand over the back of the animal and proceed to “scruff” the mouse as described above.

2. To collect urine, hold the mouse over the collection device (e.g. tube, weigh boat, aluminum foil etc.) and lightly stroke the belly of the animal. Most mice will urinate at this time. Urine is collected into the tube or onto a test strip.

Method 2:
1. Restrain animal as described in method 1.

2. Use the thumb and index finger to exert gentle, constant (30 second) pressure on the bladder until it releases.

3. To collect urine, hold the mouse over the collection device (e.g. tube, weigh boat, aluminum foil etc.)

Method 3:
1. Place the mouse in a clean, dry, empty cage or on a tabletop cover with a non-absorbable, sanitizable material (i.e. plastic wrap).

2. Animal will roam around. Animal is returned to its home cage as soon as it urinates.

3. Aspirate voided urine using a pipette and transfer to a collection tube.
NOTE: Samples collected in this manner will not be as clean as those collected during methods 1 and 2.

**Method 4:**

1. Place the animal in a metabolic cage. Animal is returned to its home cage as soon as it urinates. (Food and water must be provided if using this method.)

2. Transfer voided urine into the collection tube.

   NOTE: Samples collected in this manner will not be as clean as those collected during methods 1 and 2.

**2. Anesthetic/Analgesic Regimen**

   a. Please list all anesthetics/analgesics used in this procedure in the following table.

   If not applicable, please check here ✗ NA

   *Example:*

<table>
<thead>
<tr>
<th>Anesthetic Agent</th>
<th>Diluents Used</th>
<th>Dose &amp; Route of Administration (e.g. 1mg/kg I.V.)</th>
<th>Volume</th>
</tr>
</thead>
<tbody>
<tr>
<td>Isoflurane</td>
<td>Oxygen</td>
<td>Inhalation to effect ~2%</td>
<td></td>
</tr>
<tr>
<td>OR Tribromoethanol</td>
<td>Sterile PBS</td>
<td>400 mg/kg IP</td>
<td>0.2ml/10g body weight</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Anesthetic Agent</th>
<th>Diluents Used</th>
<th>Dose &amp; Route of Administration (e.g. 1mg/kg I.V.)</th>
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</tr>
</thead>
</table>

<table>
<thead>
<tr>
<th>Analgesic Agent</th>
<th>Dose &amp; Route of Administration (e.g. 1mg/kg I.V.)</th>
<th>Volume</th>
</tr>
</thead>
</table>

   b. Supportive care while animal recovers from anesthesia:

   N/A

**3. Post Procedure Care**

Describe post procedure care, including frequency of observations, schedule for removal of sutures/clips, etc…

N/A

**4. References if applicable:**