

# TITLE: KROMATID CHROMOSOME ANALYSIS REPORT

### I. ASSAY INFORMATION

Project Quote #	Q200401
Specimen Type	iPSC
Body Site	N/A
Sample ID	120 BB (\$005962)
Cell Line Gender	Male
Passage number (or N/A)	N/A
Study Objective	The purpose of this study is to characterize iPS cells grown <i>in vitro</i> , designated for cytogenetic analysis.

# II. CELL MAINTENANCE

Culture vessel	N/A
Media	N/A
Density (estimated)	N/A
Culture atmosphere	N/A
Culture maintenance	N/A

DocuSign Envelope ID: AEFF5DA2-ECD6-4F3A-9EF0-3037	FCA1F37D		
KromaliD	<b>Document Code:</b>	FORM-0068A	Docum
NUIIAIID			
	D * *		D

Date Effective:08-27-21

 Culture<br/>Maintenance<br/>Process<br/>Description
 N/A

 Maintenance<br/>Process<br/>Description
 N/A

**Revision:** 3.0

### **III.** CULTURE HARVEST

Culture Harvest Process Description	N/A
	Analyst Initial/Date: N/A

Material	Usage information
Harvest materials (trypsin, EDTA, etc.)	Type: N/a LN/ Exp. Date: N/a
Colcemid	LN/ Exp. Date: N/a Concentration: 0.1 µg/ml (10 µl/ml) Incubation time: N/a
Hypotonic	LN/ Exp. Date: N/a Solution: N/a Incubation time:
Fixative	Prepared Fresh, day-of-use



# IV. STAINING

Solution Type	Lot#	Exp. Date	Solution Type	Lot#	Exp. Date
Isoton II Diluent	4710610	07/12/22	Wright Stain	210817-Wright	08/17/22
Pancreatin	SLCD9444	01/11/23	Gurr Buffer	211007-Gurr	11/7/21
FBS	19J079	11/4/22	Permount	210201-01	02/1/23

Process Description	A sample of fixed cells labeled 120 BB (KromaTiD Sample ID S005962) was received at KromaTiD on 10-20-21. The fixed cells were washed twice with fixative (prepared fresh day-of-use) and the O.D. was adjusted. Drops of the final cell suspension were placed on clean slides and aged for 60 minutes at 90°C. Slides were digested in a pancreatin solution with Isoton II diluent. The enzymatic reaction was then stopped by rinsing with FBS, followed by application of a stain solution (3:1 Wright/Gurr buffer) which was poured on the slides so that it covered the entire surface. After staining for up to 1 minute, slides were washed with de- ionized water for 1-5 seconds and air dried. The mounting medium Permount was applied to the slides, a coverslip was placed on the slide and the slides were scanned on the microscope.
	Analyst Initial/Date: MV 10-26-21 through 10-29-21

# **TEST DESCRIPTION:**

G-banding with trypsin treatment and Giemsa stain (GTG-banding) is used in cytogenetics to produce a visible karyotype by staining metaphase chromosomes. This technique allows each chromosome to be distinguished by its characteristic banding pattern. G-banding is useful in assessing structural abnormalities in individual chromosomes, as well as extra or missing chromosomes within a cell. Industry-standard protocols for scoring and describing results were used (ISCN 2016: An International System for Human Cytogenomic Nomenclature).

KromaliD

### V. **RESULTS**

Cells Counted	20	Total Karyograms	2
Cells Analyzed	20	Average Band Resolution	375
Image File Location		Jax Gbanding_S005962	

#### 5.1 Chromosome Count per 20 Metaphases

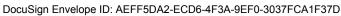
Of the 20 cells counted, 20 contained 46 chromosomes (100%). *Cells containing greater than* 57 *chromosomes are recorded as polyploid*. The polyploid frequency was 0%, based on the metaphases counted.

### 5.2 CHROMOSOME ABERRATION DATA

The chromosome aberration data via G-band for the 20 metaphases examined is summarized in attached case report cell list. 0 chromosome aberrations were found in the 20 cells analyzed with 0% of the cells aberrant.

\*Note: Cells with an euploidy gain/loss were found to be non-clonal, and therefore not included in the aberration data below.

	Tech Summary	Additional Comments		
Karyotype	46,XY[20]	Normal Male Karyotype		
Cells Analyzed	20			
Normal Cells	20	Random loss/gain cells normalized		
Abnormal Cells	0			
Aberration	n/a			
Туре				
Aberration %	0%			





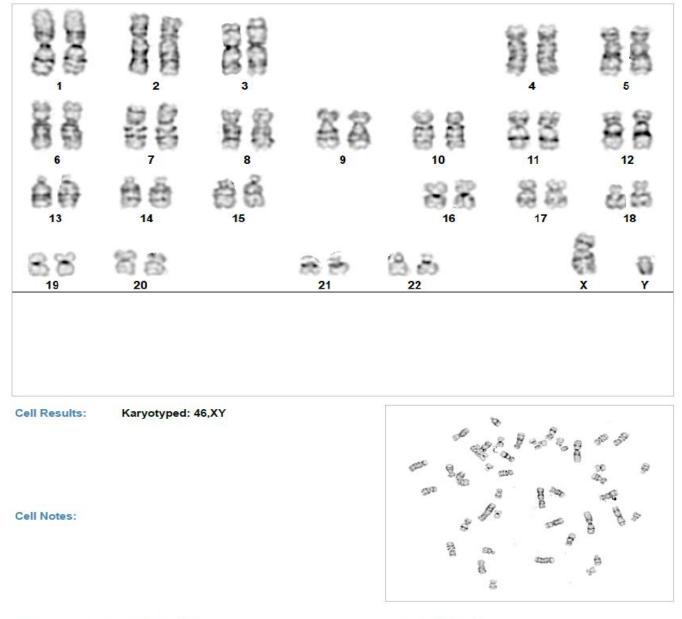
57	I CATI 37D			
	<b>Document Code:</b>	FORM-0068A	<b>Document Type:</b>	FORM
	<b>Revision:</b>	3.0	Date Effective:	08-27-21

5.3. INTERPRETATION/ SIGNIFICANCE:

G-banded chromosome analysis of metaphase cells designated as 120 BB (KromaTiD Sample ID S005962) show a normal male karyotype 46,XY[20].

The other abnormalities/aberrations detected were non-clonal, and were designated as low level mosaicism or random gain/loss.

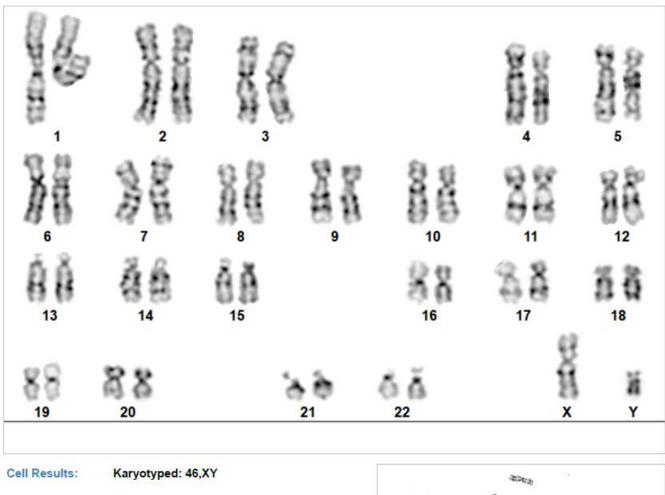




Label - Slide/Cell: S005962 - 1/27

X,Y: 15.8 , 24.6

DocuSign Envelope ID: AEFF5DA2-ECD6-4F3A-9EF0-3037FCA1F37D					
KromaliD	Document Code:	FORM-0068A	<b>Document Type:</b>	FORM	
NUIIIAIID					
	Revision:	3.0	Date Effective:	08-27-21	







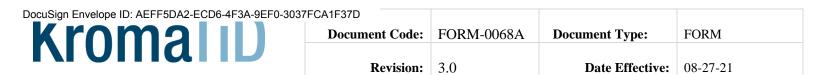
Label - Slide/Cell: S005962 - 1/31

X,Y: 17.5 , 26.1

DocuSign Envelope ID: AEFF5DA2-ECD6-4F3A-9EF0-3037FCA1F37D					
Kromali	<b>Document Code:</b>	FORM-0068A	<b>Document Type:</b>	FORM	
NUIIAID					
	<b>Revision:</b>	3.0	Date Effective:	08-27-21	

**Limitations:** This assay allows for microscopic visualization of numerical and structural chromosome abnormalities. The size of structural abnormality that can be detected is >3-10Mb, dependent upon the G-band resolution obtained from this specimen. For the purposes of this report, band level is defined as the number of G-bands per haploid genome. Detection of heterogeneity of clonal cell populations in this specimen is limited by the number of metaphase cells analyzed, documented above as "number of cells counted". Results are for Research Use Only, and should not be used for clinical purposes.

	DocuSigned by:
Completed By/Date:	Michael Vithill <sup>2021</sup>
	Michael Vernich
(	Cytogenetics Associate
	DocuSigned by:
<b>Reviewed By/ Date:</b>	Ivan fents 11/11/2021
nevieweu Dy, Duter	Ivan Perez
(	Cytogenetics Technologist III
(	$\sim$ DocuSigned by: $11/11/2021$
Approved By/Date:	DocuSigned by: Maussa Jaluguus
	5FAF272CE1894D0
	Marissa Rodrigues
	QA Manager



APPENDIX A: CASE REPORT FOR SAMPLE S005962

DocuSign Envelope ID: AEFF5DA2-ECD6-4F3A-9EF0-3037	FCA1F37D			
KromaliD	<b>Document Code:</b>	FORM-0068A	<b>Document Type:</b>	FORM
NUIIAID				
	<b>Revision:</b>	3.0	Date Effective:	08-27-21

# Table A-1: Chromosome Analysis for sample 120 BB (S005962). 20 cells were analyzed.

#	Slide	Cell	Coordinates	Results	Analysis State	State By
Slide: N	ame: 1 Labo	el: S005962				
1	1	1	16.20 X 9.54	Karyotyped: 46,XY	Karyotyped mv	ernich
2	1	15	12.03 X 20.02	Karyotyped: 46,XY	Karyotyped mv	ernich
3	1	16	14.52 X 19.29	Karyotyped: 46,XY	Karyotyped mv	ernich
4	1	19	15.98 X 19.57	Karyotyped: 46,XY	Karyotyped mv	ernich
5	1	26	17.99 X 22.89	Karyotyped: 46,XY	Karyotyped mv	ernich
6	1	27	15.77 X 24.64	Karyotyped: 46,XY	Karyotyped mv	ernich
7	1	31	17.55 X 26.10	Karyotyped: 46,XY	Karyotyped mv	ernich
8	1	33	15.72 X 26.59	Karyotyped: 46,XY	Karyotyped mv	ernich
9	1	41	9.45 X 29.73	Karyotyped: 46,XY	Karyotyped mv	ernich
10	1	42	10.44 X 31.34	Karyotyped: 46,XY	Karyotyped mv	ernich
11	1	43	12.66 X 30.53	Karyotyped: 46,XY	Karyotyped mv	ernich
12	1	45	17.58 X 31.10	Karyotyped: 46,XY	Karyotyped mv	ernich
13	1	48	8.55 X 32.86	Karyotyped: 46,XY	Karyotyped mv	ernich
Slide: N	ame: 2 Labe	el: S005962				
14	2	2	11.02 X 14.66	Karyotyped: 46,XY	Karyotyped mv	ernich
15	2	3	17.97 X 14.93	Karyotyped: 46,XY	Karyotyped mv	ernich
16	2	12	15.10 X 19.37	Karyotyped: 46,XY	Karyotyped mv	ernich
17	2	14	16.73 X 21.93	Karyotyped: 46,XY	Karyotyped mv	ernich
18	2	28	9.95 X 34.25	Karyotyped: 46,XY	Karyotyped mv	ernich
19	2	32	12.32 X 35.59	Karyotyped: 46,XY	Karyotyped mv	ernich
20	2	37	14.88 X 41.24	Karyotyped: 46,XY	Karyotyped mv	ernich